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Tips and tricks for optimizing sperm preparation and Minimizing DNA fragmentation in the andrology process

The integrity of sperm DNA is essential for healthy embryo development as chromatin abnormalities or DNA damage may result in lower fertilization rates in IVF, impaired implantation rates and an increased incidence of abortion¹. Sperm DNA fragmentation can be caused by intrinsic factors but also occurs as a result of extrinsic factors including storage temperatures, handling conditions, and time between ejaculation and preparation among others. Appropriate semen processing is critical for successful infertility treatment, therefore **ORIGIO has worked with leading experts to compile the following tips and tricks to optimize sperm preparation and minimize DNA fragmentation during sperm processing.**

Abstinence period:

It has been suggested that the man abstains from sexual activity for just one day before sample production for treatment as this has been shown to reduce DNA fragmentation ^{2, 3}. Patients might also be encouraged to ejaculate frequently prior to treatment so that spermatozoa spend the minimum period of time within the epididymis, generally accepted to be the site where DNA damage occurs *in vivo*.

Time between ejaculation and sample preparation:

As DNA fragmentation is known to increase, it is recommended that the time between sample production and the start of preparation is kept as short as possible and so should begin immediately after full liquefaction. If a sample must be produced at home, it should ideally be delivered to the laboratory within one hour and should be kept at 15-25°C; it should not be allowed to get too cold or too warm.

Viscous semen samples

To obtain satisfactory sperm yield from highly viscous semen samples it is recommended to dilute the viscous sperm sample 1:1 with a sperm washing medium (e.g. ORIGIO Sperm Preparation Medium or SAGE Quinn's Sperm Washing Medium) followed by repeated pipetting¹⁹.

Sperm DNA fragmentation assay:

Sperm DNA is not evaluated by a standard semen analysis but can provide essential information which can be used to guide couples to treatment that is appropriate for their particular needs.

Sperm preparation:

It is advised to use media containing anti-oxidants (*e.g. taurine, EDTA, citrate and HSA*) as oxidative stress is known to be the main mechanism causing DNA fragmentation¹⁷.

Simple Sperm washing:

Simple washing should be avoided whenever possible since damage can be caused by reactive oxygen species (ROS) generated by non-viable spermatozoa and leukocytes. Additionally, the presence of large numbers of non-viable spermatozoa in the prepared sample can inhibit capacitation⁴.

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Gradient preparation and Swim-Up:

Rates of DNA damage are decreased by both swim-up and density gradient centrifugation, though it has been suggested that the gradient technique gives more stable sperm in terms of rate of DNA fragmentation⁵. Motile sperm obtained by density gradient centrifugation are shown to have a higher mitochondrial membrane potential and a lower rate of DNA fragmentation, to generate lower levels of ROS, and are more viable than those in whole semen⁶.

Centrifugation time:

This should be kept as short as possible (15-20 min) in order to minimize the production of ROS by leukocytes and non-viable sperm cells. Longer centrifugation time also increases the temperature and affects the quality.⁶

Number of washes after density gradient:

It is important to wash samples twice following density gradient centrifugation; this removes any colloidal particles and has been shown to give a higher yield of rapidly progressive sperm⁷.

Storage and temperature:

Sperm DNA damage is enhanced by prolonged incubation, especially at 37°C. The DNA fragmentation rate at room temperature is found to be significantly lower than that at 37°C after 24 h⁸. Handling and preparation should therefore always take place at room temperature, preferably in the dark, and for the shortest amount of time before insemination or use in treatment.

Sperm preparation for HIV positive men:

It is recommended to add an additional swim-up step of sperm from the pellet resulting from the density gradient. The swim-up step into clean media further separates the sperm from any leukocytes in the pellet, resulting in a cleaner sperm sample for insemination ²⁰.

Zika Virus - precautions

Men with possible exposure to the Zika virus but without clinical illness consistent with Zika virus disease should wait at least 8 weeks after exposure to attempt conception. At present no data or recommendations are available for purifying sperm away from these viruses in semen; all counselling, precautions and procedures recommended for other viruses should be followed if there is any suspicion of infection²⁰.

Choose the most motile sperm for IVF and ICSI:

Choose the most motile fraction of a prepared semen sample for fertilization since an inverse correlation is found between sperm motility and the degree of sperm DNA fragmentation in patients with clinical varicocele and other conditions¹³.

IVF vs ICSI in men with high sperm DNA damage:

Reports¹⁶ show a decreased live birth rate after IVF for men with high sperm DNA fragmentation, whilst live birth rates are unaffected when ICSI is used. It may be preferable to perform ICSI for men with high sperm DNA fragmentation, though this needs more investigation¹⁶.



Sperm Selection for ICSI:

It is known that apparently morphologically normal sperm from men with poor semen quality are at higher risk of being aneuploid¹⁰ so the usual approach of choosing sperm for injection using motility and morphology is not completely effective. The additional use of hyaluronan binding is known to reduce the risk of selecting a sperm with fragmented DNA and improve embryo quality¹¹ and has been shown to reduce the risk of early pregnancy loss¹².

Cryopreservation:

Timing: Frozen-thawed sperm exhibit different dynamics of DNA fragmentation compared with fresh samples with a more rapid increase in the percentage of DNA damaged spermatozoa¹⁴. If cryopreservation is required, it is recommended that sperm be used in treatment as soon as possible after thawing.

Osmotic shock: The osmotic shock phenomenon is characterized by the increased coiling of the sperm tail and can results in loss of progressive motility. It is therefore important to allow for gradual osmotic adjustment by slowly mixing the Sperm Freezing medium with your sperm sample.

References:

- 1. Simon L *et al* (2014) Comparative analysis of three sperm DNA damage assays and sperm nuclear protein content in couples undergoing assisted reproduction treatment. *Hum Reprod*, **29**: 904–917, 2014
- 2. Pons I *et al* (2013) One day abstinence decreases sperm DNA fragmentation in 90% of selected patients. *J Assist Reprod Genet* **30**:1211-1218
- 3. Gosalvez J *et al* (2011). Shorter abstinence decreases sperm deoxyribonucleic acid fragmentation in ejaculate. *Fertil Steril* **96**:1083–6.
- 4. Beydolaet T *et al* (2014). Sperm Preparation and Selection Techniques. Medical and surgical management of male infertility. *New Delhi; Philadelphia : Jaypee Brothers Medical Publishers*. Section 29
- 5. Zhang X-D et al (2011). The effects of different sperm preparation methods and incubation time on the sperm DNA fragmentation. *Hum Fertil* **14**: 187-191.
- 6. Ashok D (2014). Strategies to Ameliorate Oxidative Stress during Assisted Reproduction. Springer ISSN 2194-4253
- 7. Turhan N *et al.*, (2011). Single or double sperm wash processing by density gradient centrifugation: effect on clomiphene citrate induced intrauterine insemination cycle outcomes. *Turk J Med Sci.* **41** (1): 39-44
- 8. Matsuura R *et al.* (2010) Preparation and incubation conditions affect the DNA integrity of ejaculated human spermatozoa. *Asian J Androl* **12**: 753–759
- 9. Ghumman S et al., (2011) Combination of swim-up and density gradient separation methods effectively eliminate DNA damaged sperm. J Turkish-German Gynecol Assoc 12: 148-52
- 10. Burello N *et al* (2004) Morphologically normal spermatozoa of patients with secretory oligo-astheno-teratozoospermia have an increased aneuploidy rate. *Hum Reprod* **19**: 2298-2302
- 11. Parmegiani *et al* (2010) Physiologic ICSI": hyaluronic acid (HA) favors selection of spermatozoa without DNA fragmentation and with normal nucleus, resulting in improvement of embryo quality. *Ferti Steril* **93**:598-604.
- Worrilow K et al (2013) Use of hyaluronan in the selection of sperm for intracytoplasmic sperm injection (ICSI): significant improvement in clinical outcomes—multicenter, doubleblinded and randomized controlled trial Hum Reprod 28:306–314
- 13. Peluso G *et al* (2013). The study of spermatic DNA fragmentation and sperm motility in infertile subjects *J Androl;* 27: 106–111
- 14. Gozalves J *et al* (2009) A dynamic assessment of sperm DNA fragmentation versus sperm viability in proven fertile human donors. *Ferti Steril* **92**:1915–9
- 15. Simon L *et al* (2013) Sperm DNA damage has a negative association with live-birth rates after IVF. *Reprod BioMed Online* **26**, 68–78
- 16. Osman A *et al* (2015) The effect of sperm DNA fragmentation on live birth rate after IVF or ICSI: a systematic review and meta-analysis *Reprod BioMed Online* **30**, 120–127
- 17. Amrit B et al (2010). Impact of oxidative stress and antioxidants on semen function. Vet Med Int. 2011; pii: 686137.
- 18. Said T. M. et al (2010). Implication of apoptosis in sperm cryoinjury. Reprod BioMed. 21, no. 4, pp. 456–462.
- 19. WHO laboratory manual for the Examination and processing of human semen. (2010) 5th edition. Part 1, section 2.3.1.1

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20. Jindal SK *et al* (2016). Guidelines for risk reduction when handling gametes from infectious patients seeking assisted reproductive technologies. *Reprod Biomed Online*. Aug; **33** (2):121-30.